Chlorothalonil ELISA Kit

*Cat.No: DEIANJ23
Lot. No. (See product label)*

**Size**

96T

**Intended use**

The Chlorothalonil ELISA is an immunoassay for the detection of chlorothalonil in contaminated samples.

**General Description**

Chlorothalonil is an organic compound mainly used as a broad spectrum, nonsystemic fungicide, with other uses as a wood protectant, pesticide, acaricide, and to control mold, mildew, bacteria, algae.

**Principle Of The Test**

The test is a direct competitive ELISA based on the recognition of Chlorothalonil by specific antibodies. Chlorothalonil, when present in a sample, and a Chlorothalonil-enzyme conjugate compete for the binding sites of anti-Chlorothalonil antibodies in solution. The Chlorothalonil antibodies are then bound by a second antibody immobilized on the plate. After a washing step and addition of the substrate solution, a color signal is produced. The intensity of the blue color is inversely proportional to the concentration of Chlorothalonil present in the sample. The color reaction is stopped after a specified time and the color is evaluated using a microplate ELISA photometer. The concentrations of the samples are determined by interpolation using the standard curve constructed with each run.

**Reagents And Materials Provided**

1. Microtiter plate coated with a second antibody.
2. Chlorothalonil Standards.
3. Chlorothalonil-HRP Conjugate.
5. Anti-Chlorothalonil Antibody Solution.
7. Wash Solution.
8. Color (Substrate) Solution (TMB).
9. Stop Solution.

**Materials Required But Not Supplied**

1. Micro-pipettes with disposable plastic tips (20-200 µL)
2. Multi-channel pipette or stepper pipette (50-250 µL) with disposable plastic tips
3. Deionized or distilled water
4. Graduated cylinder
5. Container with 500 mL capacity (for 1X diluted Wash Solution)
6. Tape or Parafilm
7. Timer
8. Paper towels or equivalent absorbent material
9. Microtiter plate shaker (optional)
10. Microtiter plate washer (optional)
11. Microtiter plate reader (wave length 450 nm)

**Storage**

Store the kit at 2 - 8°C.

**Assay Procedure**

1. Add 50 µL of the standard solutions and samples into the wells of the test strips according to the working scheme given. Analysis in duplicate or triplicate is recommended.
2. Add 50 µL of reconstituted enzyme conjugate solution to the individual wells successively using a multi-channel pipette or a stepping pipette.
3. Add 50 µL of antibody solution to the individual wells successively using a multi-channel pipette or a stepping pipette. Cover the wells with parafilm or tape and mix the contents by moving the strip holder in a circular motion on the benchtop for 30 seconds. Be careful not to spill the contents.
4. Incubate the strips for 60 minutes at room temperature.
5. After incubation, remove the covering and vigorously shake the contents of the wells into a sink. Wash the strips five times using the 1× washing buffer solution. Use at least a volume of 250 µL of washing buffer for each well and each washing step. Remaining buffer in the wells should be removed by patting the plate dry on a stack of paper towels.
6. Add 100 µL of substrate (color) solution to the wells using a multi-channel pipette or a stepping pipette. Cover the wells with parafilm or tape and mix the contents by moving the strip holder in a circular motion on the benchtop for 30 seconds. Incubate the strips for 30 minutes at room temperature. Protect the strips from direct sunlight.
7. Add 50 µL of stop solution to the wells using a multi-channel pipette or a stepping pipette in the same sequence as for the substrate solution.
8. Read the absorbance at 450 nm using a microplate ELISA photometer within 15 minutes after the addition of the stopping solution.